STEROIDAL GLYCOSIDE-PROTODIOSCIN FROM Digitalis ciliata

L. N. Gvazava¹ and V. S. Kukoladze²

UDC 547.918

Digitalis ciliata Trautv. is endemic to the Caucuses and widely distributed in the flora of Georgia. It has been proposed as raw material for producing an effective cardiotonic preparation of acetyldigitoxin [1]. Triterpene glycosides [2, 3], carotinoids [4], and steroidal glycosides were detected in the remaining extracts, the mother liquors.

After exhaustive extraction of cardenolide glycosides, the remaining solution was condensed to a resinous condition and subjected (5 g) to total acid hydrolysis with HCl (2 N). Chromatography of the hydrolysate over a column of Al_2O_3 using ether, ether:benzene, and benzene:chloroform isolated three genins that were identified as tigogenin, diosgenin, and gitogenin by their physical constants and TLC in the presence of authentic samples.

The insoluble solid that contained mainly spirostane glycosides was separated. The aqueous phase was extracted several times with n-butanol. The extracts were combined and evaporated to dryness. The solid was dissolved in ethanol. The glycosides were precipitated with acetone to afford an amorphous cream-colored powder (12.5 g), a portion (3.5 g) of which was chromatographed successively over columns of silica gel (KSK, 40-100 μ m) and Sephadex G-75 using CHCl₃:CH₃OH:H₂O (65:35:8) to afford a mixture of glycosides 1 and 2, which gave two spots on TLC with a small difference (ΔR_f 0.1) in mobility. Rechromatography over the columns isolated the pure compounds 1 (0.185 g) and 2 (0.076 g). Both glycosides gave a positive reaction with Ehrlich's reagent [5]. Their IR spectra lacked signals characteristic of the spiroketal group. These data indicated that the compounds were furostanes.

Glycoside **1**, white amorphous powder (ethanol), mp 189-192°C, $[\alpha]_D^{25}$ -80.5° (CHCl₃:CH₃OH, 1:1, c 0.5), lit. [6] mp 190-196°C, $[\alpha]_D^{25}$ -79.8° (pyridine, c 0.1). IR spectrum (KBr, v, cm⁻¹): 3400 (OH), 1045, 930 (weak broad band), 915, 840. FAB MS (m/z, %): 1071 [M + Na]⁺, 925 [M + Na - deoxyhexose]⁺, 891 [M + Na - hexose - H₂O]⁺.

The acid hydrolysate of **1** contained diosgenin. GC of the aldonitriles of the sugars identified glucose and rhamnose in a 1:1 ratio. Hakomori [7] methylation of **1** and subsequent methanolysis produced in the hydrolysate 2,3,4-tri-*O*-methyl-L-rhamnose, 2,3,4,6-tetra-*O*-methyl-D-glucose, and 3,6-di-*O*-methyl-D-glucose.

Enzymatic hydrolysis of **1** by β -glucosidase gave the prosapogenin that was identified using physical constants and mass and NMR spectra as dioscin [8]. The hydrolysate contained D-glucose. Considering these data, **1** was assigned the structure 3-O-{[α -L-rhamnopyranosyl-(1 \rightarrow 4)]-[α -L-rhamnopyranosyl-(1 \rightarrow 2)]- β -D-glucopyranosyl}-(25R)-furost-5-en-3 β ,22 α ,26-triol-26- β -D-glucopyranoside or protodioscin [9].

This glycoside was isolated from plants of the genus *Digitalis* for the first time.

Glycoside **2** had molecular weight m/z 1085 [M + Na]⁺ according to FAB MS, 14 amu greater than that of **1**. The PMR spectrum had a 3H singlet at δ 3.26 (OMe). Thus, it was assumed that **2** was the 22-*O*-methyl ether of **1**, which was consistent with its PMR and ¹³C NMR spectra and agreed well with the literature [10] (Table 1).

¹⁾ I. Kutateladze Institute of Pharmaceutical Chemistry, 0159, Tbilisi, e-mail: liligvazava@yahoo.com; 2) P. Melikishvili Institute of Physical and Organic Chemistry, 0186, Tbilisi, ul. Dzhikia, 5. Translated from Khimiya Prirodnykh Soedinenii, No. 5, pp. 495-496, September-October, 2006. Original article submitted August 18, 2006.

TABLE 1. Chemical Shifts in PMR and 13 C NMR Spectra of 2 (δ , ppm, J/Hz, pyridine-d₅, 0 = HMDS)

C atom	δ_{C}	C atom	δ_{C}	Proton	δ_{H}
1	37.5	D-Glucose		Me-18	0.84 (s)
2	29.8	1'	101.4*	Me-19	1.04 (s)
3	77.6	2'	77.9	Me-21	1.17 (d, J = 7.4)
4	39.3	3'	74.7	Me-27	1.00 (d, J = 7.4)
5	140.9	4 ′	77.4	CH2-26	3.62 (br.d, $J = 11.8$)
6	121.9	5 ′	75.9	H-6	5.32 (br.d, $J = 9.1$)
7	32.2	6'	61.9	H-1'	4.88 (d, J = 7.7)
8	31.6	L-Rhamnose		H-1"	6.28 (br.s)
9	50.3	1"	99.3	H-1'''	5.80 (br.s)
10	36.9	2"	72.5	H-1""	4.78 (d, J = 7.1)
11	21.1	3"	71.4	Me"	1.71 (d, J = 6.0)
12	40.0	4 "	75.9	Me'''	1.60 (d, J = 6.0)
13	40.7	5"	69.3	OMe	3.26 (s)
14	56.5	6 "	18.5		
15	32.4	L-Rhamnose			
16	81.1	1‴	103.5*		
17	64.0	2′′′	72.6		
18	16.3	3‴	71.6		
19	19.5	4 ′′′	75.9		
20	41.8	5‴	68.6		
21	16.5	6 '''	18.4		
22	112.8	D-Glucose			
23	30.5	1""	103.7		
24	28.1	2""	74.8		
25	33.7	3""	77.5		
26	74.8	4 ""	71.4		
27	17.3	5""	77.5		
		6 ""	63.6		

REFERENCES

- 1. L. N. Gvazava and E. P. Kemertelidze, USSR Pat. No. 611427; Byull. Izobret., No. 2394451, 3 (1978).
- 2. E. P. Kemertelidze, L. N. Gvazava, M. D. Alania, V. S. Kikoladze, and V. G. Tsitsishvili, *J. Nat. Prod.*, **55**, 217 (1992).
- 3. E. P. Kemertelidze, L. N. Gvazava, M. D. Alaniya, and V. S. Kikoladze, *Khim. Prir. Soedin.*, 246 (1991).
- 4. L. N. Gvazava, *Izv. Akad. Nauk Gruz.*, *Ser. Khim.*, **16**, No. 4, 317 (1990).
- 5. S. Kiyosava, N. Huton, T. Komori, T. Nohara, J. Hosokava, and T. Kavasaki, *Chem. Pharm. Bull.*, **16**, 1162 (1968).
- 6. T. Kawasaki, T. Komori, and K. Miyahara, *Chem. Pharm. Bull.*, **22**, 2164 (1974).
- 7. S. Hakomori, J. Biochem. (Tokyo), **55**, 205 (1964).
- 8. T. Kawasaki, H. Yabuta, M. Suenobu, et al., *Chem. Pharm. Bull.*, **21**, 1240 (1973).
- 9. T. Nohara, F. Kumamoto, K. Miyahara, and T. Kawasaki, Chem. Pharm. Bull., 23, 1158 (1975).
- 10. P. K. Agrawal, D. C. Jain, R. K. Gupta, and R. S. Thakur, *Phytochemistry*, 24, 2479 (1985).